



Is the Future of Phenotypic-based Identification of Biothreat Agents in Jeopardy of the Rule Out or Refer Practice?

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OBJECTIVES

- Identify three reasons that contributed to decline in utilization of the Sentinel Level Protocols in support of the Rule Out/Refer Concept
- What is the cost comparison between conventional rule out tests versus newer technologies?
- Identify the primary training resource that nonpublic health clinical laboratories rely on for training related to biothreat and emerging infectious disease agents



Contributing Factors to the Decline in the use of LRN Sentinel Level Protocols

- **New Technology and Personnel Shortages**

- Mass Spectrometry (MALDI-TOF)
- Some media not being retained in laboratory inventory (e.g. Urea, Motility)
- Lack of well trained, skilled laboratory scientists
“the dumbing down process”



Shortage of Trained Personnel

Administrators Belief that Automation is the Final Solution

- Aging (Avg. age: 56 years)
- Clinical Laboratory Science Programs:
- **not teaching basic concepts/fundamentals**
 - clinical laboratory rotation relied upon to prepare students for microbiology
 - students not exposed to organism culture characteristics and Gram stain morphology
 - “you will have an automated or conventional system making the ID for you”
 - accepting the ID as 100% correct without question
 - little or no training of Biothreat Agents
 - 18 months or more of training to work independently



Media and Reagents Required in Conventional Rule Out/Refer Protocols

Media

- Blood, MacConkey, Chocolate, Urea, Motility

Reagents

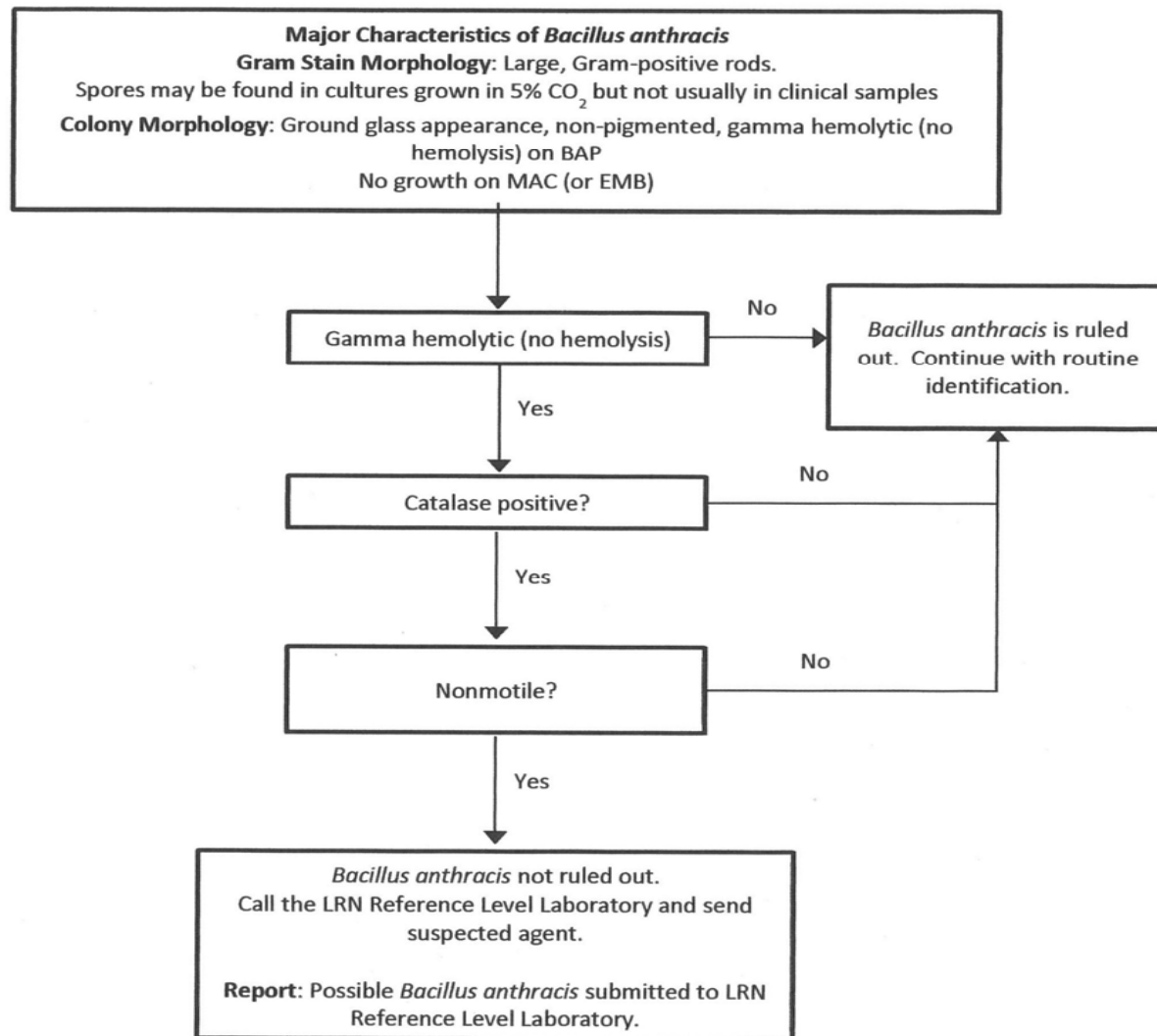
- Gram stain, Catalase, Oxidase, Indole, Cefinase, *Staphylococcus aureus* ATCC 25923

Antibiotic Disks

- Colistin, Amoxicillin/Clavulanate, Penicillin



Bacillus anthracis Identification Flowchart



Note: Biochemical test procedures and quality control instructions can be found at the end of the General Recommendation and Biochemical Testing Procedures document.



***Brucella* Identification Flowchart**

SAFETY: As soon as *Brucella* is suspected, perform ALL further work in a Class II Biosafety Cabinet (BSL3)

Major Characteristics of *Brucella* species
Gram Stain Morphology: Small (0.4 x 0.8 μ m), Gram-negative coccobacillus

THINK BRUCELLA

Growth: Subculture positive aerobic blood culture bottle to:
BAP, CHOC
Incubate in 5-10% CO₂ at 35°C

Spot BAP with *S. aureus* ATCC 25923 for satellite test.

Note poorly growing colonies after 24 hours incubation on BAP and CHOC.
Incubate plates for at least 2 additional days if no growth in 24 hours.
Organism does not grow on MAC.

Is the organism only growing on BAP
without the need to satellite around the *S. aureus* at 24-48 hours?

No

Consider
Haemophilus

Yes

Oxidase positive and catalase
positive?

No

Think *Francisella*
(see procedure)

Yes

Urea positive?

No

Reincubate and
see written
procedure

Yes

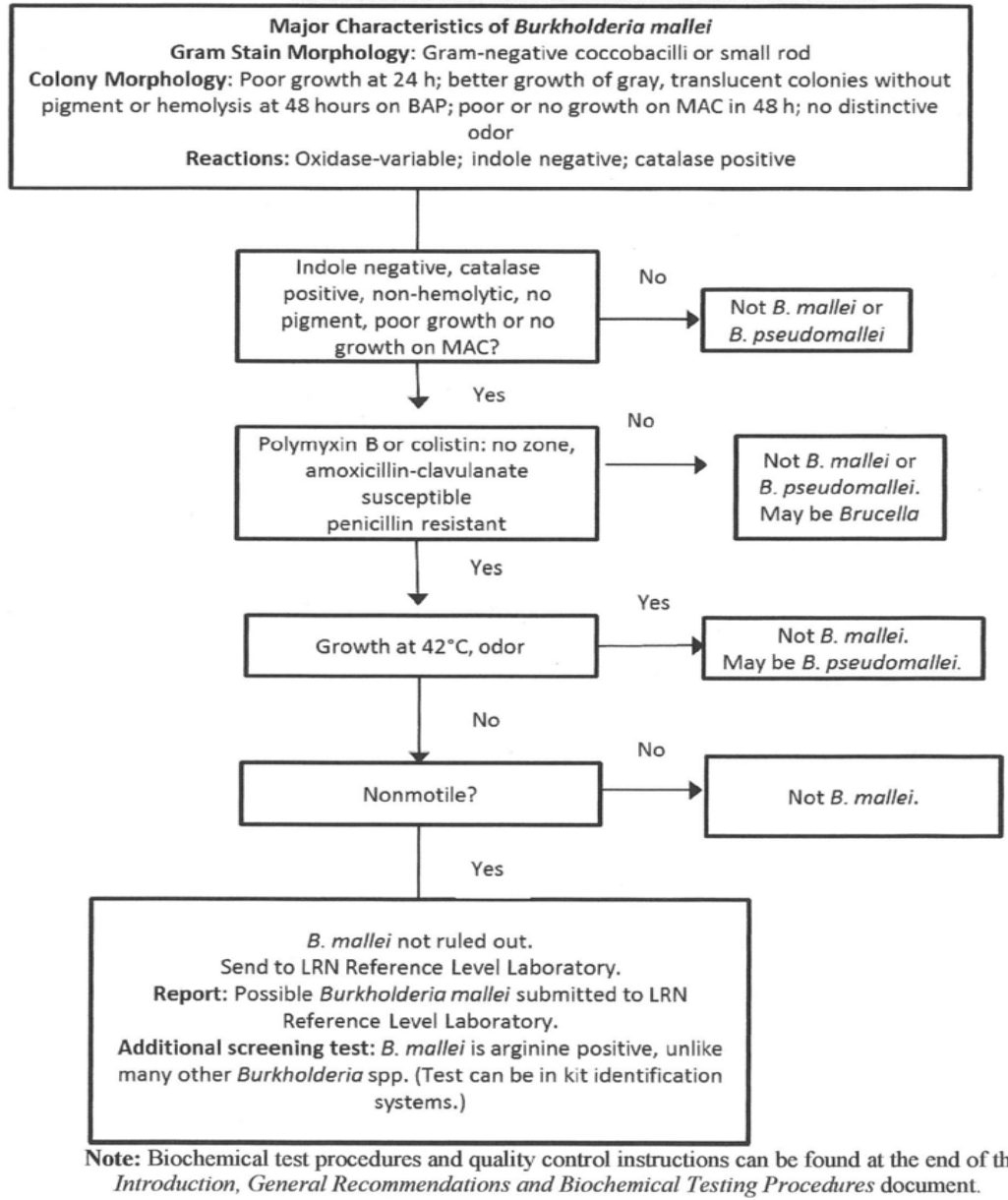
Brucella not ruled out.
Send to LRN Reference Level Laboratory.
Inform physician that *Brucella* species cannot be ruled out.

Antimicrobial therapy: Rifampin or Streptomycin plus
Doxycycline

Note: Biochemical test procedures and quality control instructions can be found at the end of the *General Recommendation and Biochemical Testing Procedures* document.



***Burkholderia mallei* Identification Flowchart**





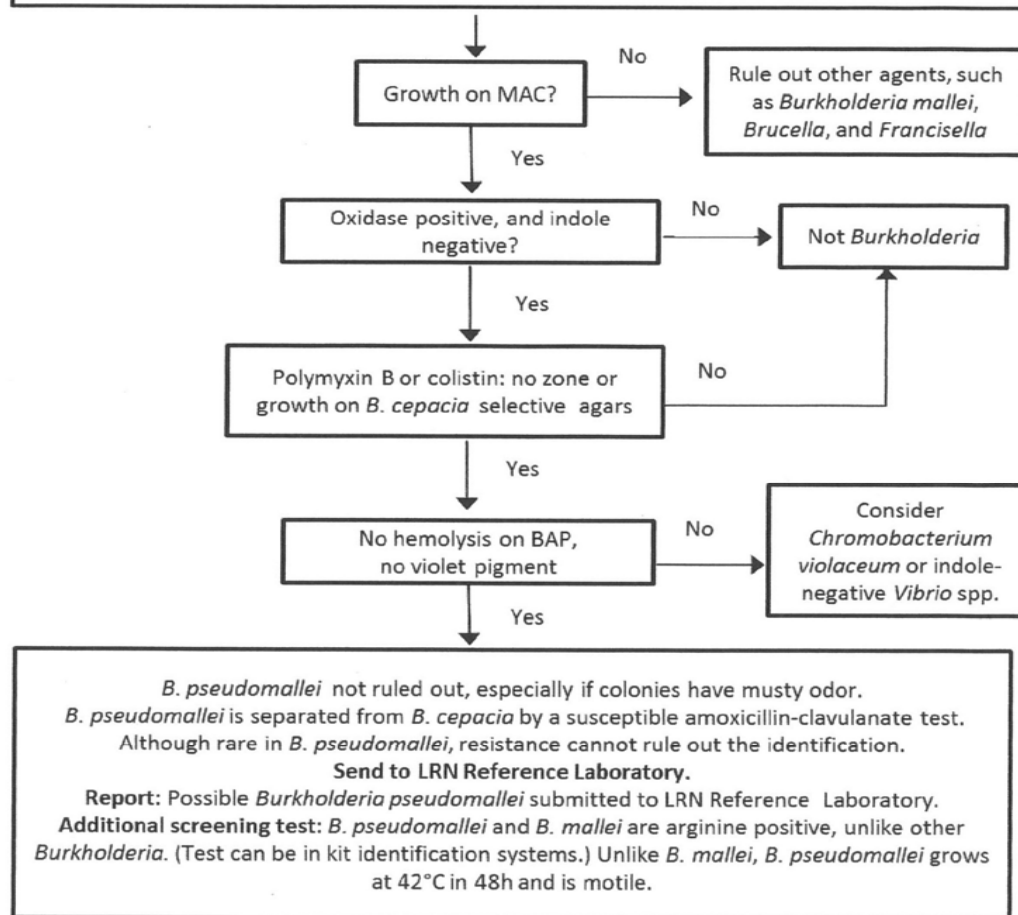
***Burkholderia pseudomallei* Identification Flowchart**

Major Characteristics of *Burkholderia pseudomallei*

Gram Stain Morphology: Gram-negative rod, straight or slightly curved, may demonstrate bipolar morphology at 24 h and peripheral staining, like endospores, as cultures age.

Colony Morphology: Poor growth at 24 h, good growth of smooth, creamy colonies at 48h on BAP, may develop wrinkled colonies in time, nonhemolytic. Can demonstrate strong characteristic musty, earthy odor, growth on MacConkey in 48 h., no pigment is visible on Mueller-Hinton agar, may have non-violet pigment on BAP.

Reactions: Oxidase-positive, indole negative



Note: Biochemical test procedures and quality control instructions can be found at the end of the *Introduction, General Recommendations and Biochemical Testing Procedures* document.



***Francisella tularensis* Identification Flowchart**

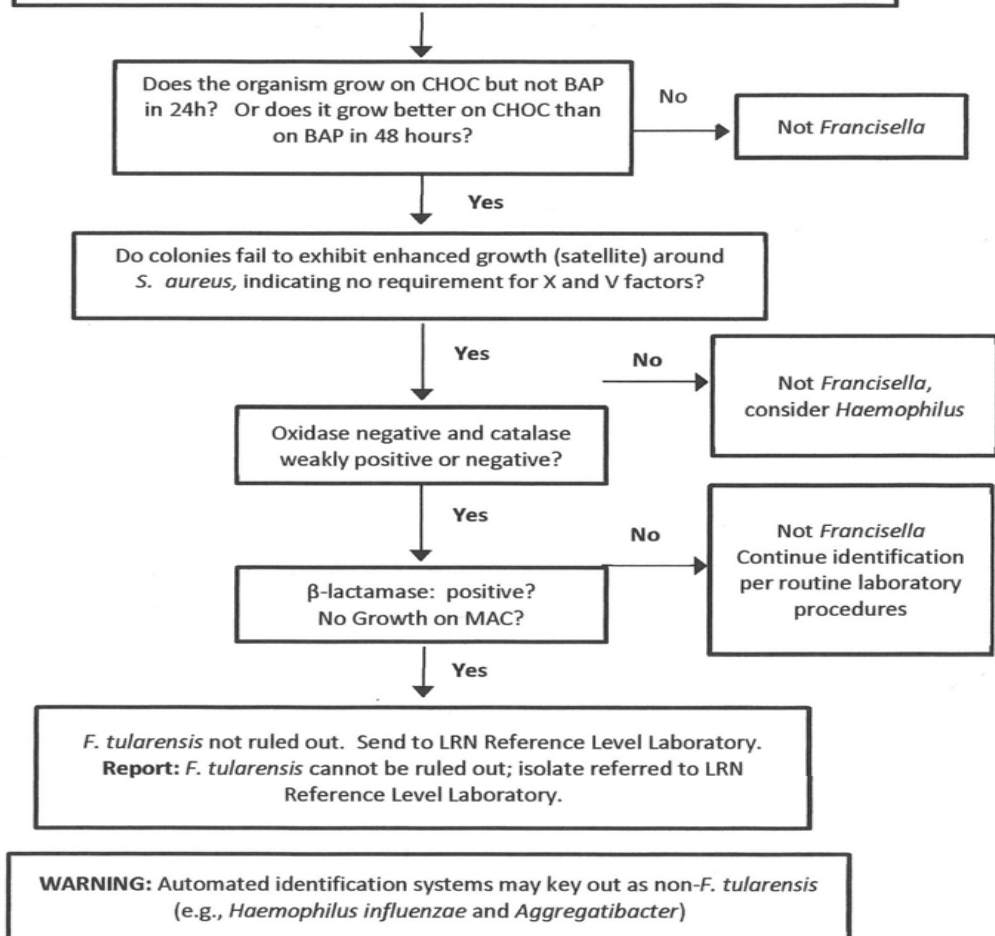
Major Characteristics of *Francisella tularensis*

Gram Stain Morphology: pleomorphic, minute (0.2 to 0.5 by 0.7 to 1.0 μm) faintly staining, Gram-negative coccobacillus.

Colony Morphology: No growth on MAC, scant to no growth on BAP after >48 h.

Produces 1-2 mm gray to grayish-white colonies on CHOC after >48 h.

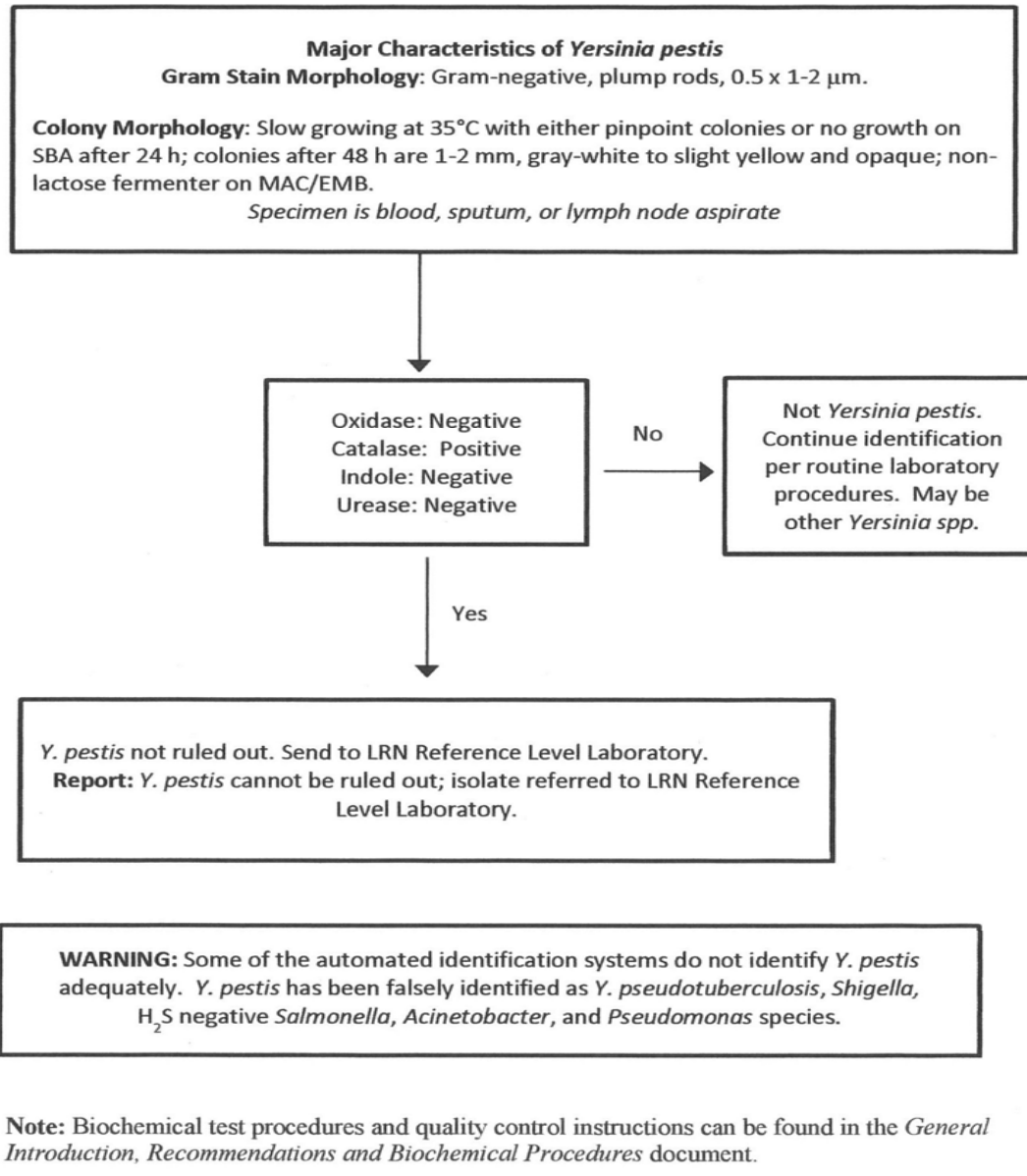
Perform all work in a biosafety cabinet using BSL-3 precautions.



Note: Biochemical test procedures and quality control instructions can be found at the end of the *General Recommendation and Biochemical Testing Procedures* document.



Yersinia pestis Identification Flowchart





Cost Comparison between Standard Rule Out and MALDI-TOF

Agent	Rule Out Protocol		MALDI Protocol	
	Reagent Cost	Labor Cost	Reagent Cost	Labor Cost
<i>Bacillus anthracis</i>	\$2.58	\$5.13	\$2.18/test*	
<i>Brucella</i>	\$1.73	\$5.64		
<i>Burkholderia mallei</i>	\$3.86	\$6.14		
<i>Burkholderia pseudomallei</i>	\$1.47	\$5.60		
<i>Francisella tularensis</i>	\$1.37	\$4.54		
<i>Yersinia pestis</i>	\$1.65	\$5.64		

*Cost includes labor, reagents, template. Does NOT include cost of instrumentation.



Issues Regarding MALDI-TOF

- **Data Bases**
 - Clinical Application (CA)
 - Research Use Only (RUO)
 - Security Software*
 - **Incorrect Identifications of Biothreat Agents**
 - *Yersinia pestis* ID as *Y. pseudotuberculosis*
 - *Bacillus anthracis* ID as *B. cereus*
 - *Burkholderia pseudomallei* ID as *B. Thailandensis*
- * Most clinical laboratories do not have Security Software



MALDI-TOF Issues

- **Lack of focus on safety in processing isolates for ID**
 - risk assessment/mitigation absent
 - spotting of isolate
 - solutions potentially a biohazard
 - effective disinfection of reusable templates
 - *Bacillus cereus* biovar *anthracis*??
 - ionization chamber/flight tube under vacuum with air expelled from the instrument; incomplete inactivation of organism is a risk for dispersal of viable bacteria via exhaust
 - most laboratories routinely perform direct colony testing, not tube-based extraction
 - presence of a hazardous bacterium is often not recognized until it has been identified

Cunningham, S. A. and R. Patel. 2015. J. Clin Microbiol. 53: 2788



The Critical Need for Training of Laboratory Personnel for Responding to BT agents

- Public Health has a major role
- Clinical Laboratory Science Programs
- Medical School
- Support for Continuing Education Programs



Summary

- Essential Media in support of rule out/refer not being maintained by the laboratory
- Reduction in skilled and qualified personnel
- New technology: faster, simple, requires less labor, and generally accurate
- MALDI: Cheaper and faster vs conventional rule out/refer methods
- Noncultural methods slowly replacing culture-based methods
- The days of phenotypic-based Rule Out/Refer are nearing an end
- Safety a major concern



NEW TECHNOLOGY HAS COME AND IS
REPLACING WHAT WE HAVE BEEN
DOING FOR A CENTURY OR MORE