

State Hygienic Laboratory

COVID-19 Wastewater Surveillance: Sampling Frequency and Hospitalization Trends

Ryan Jepson, MS

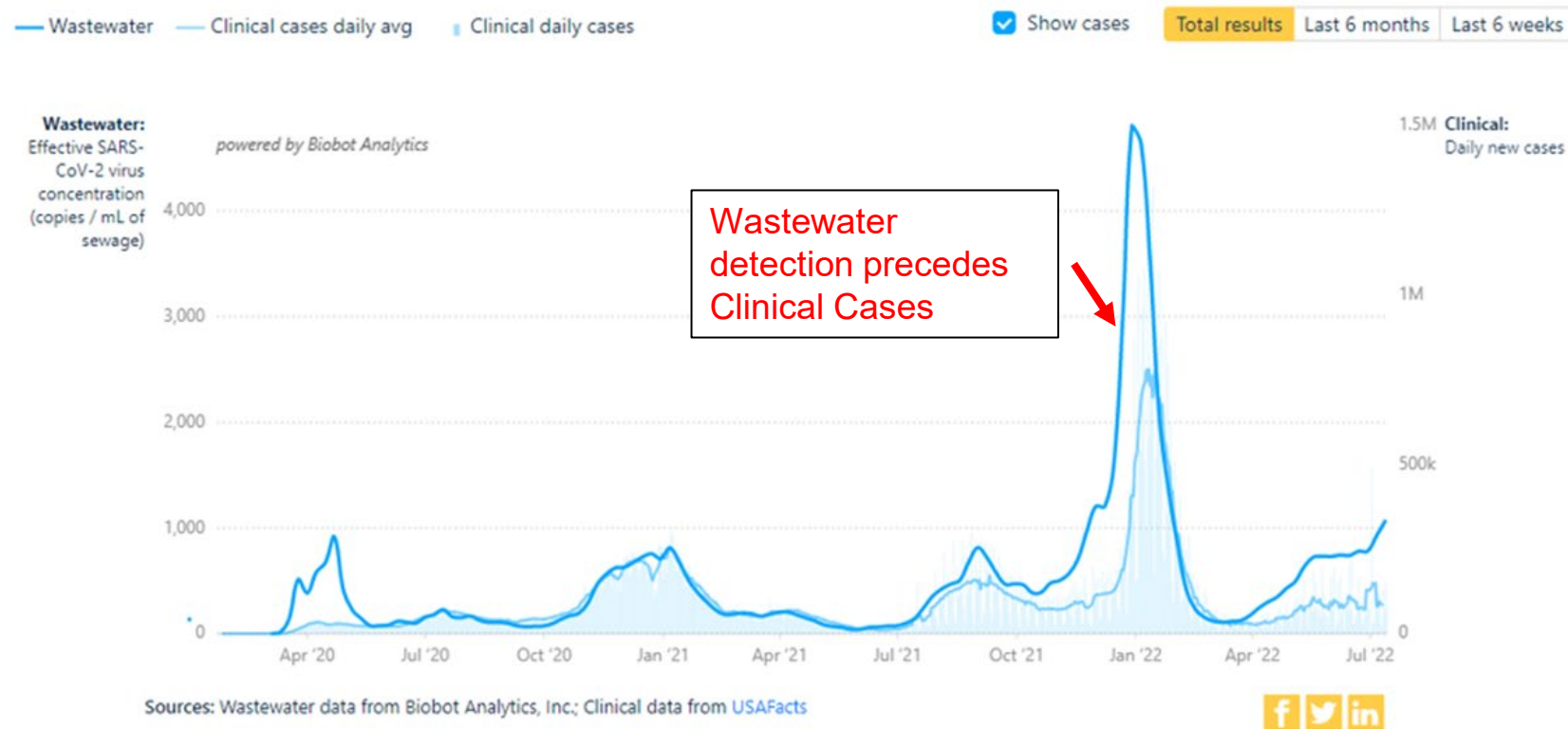
March 26, 2025

Introduction



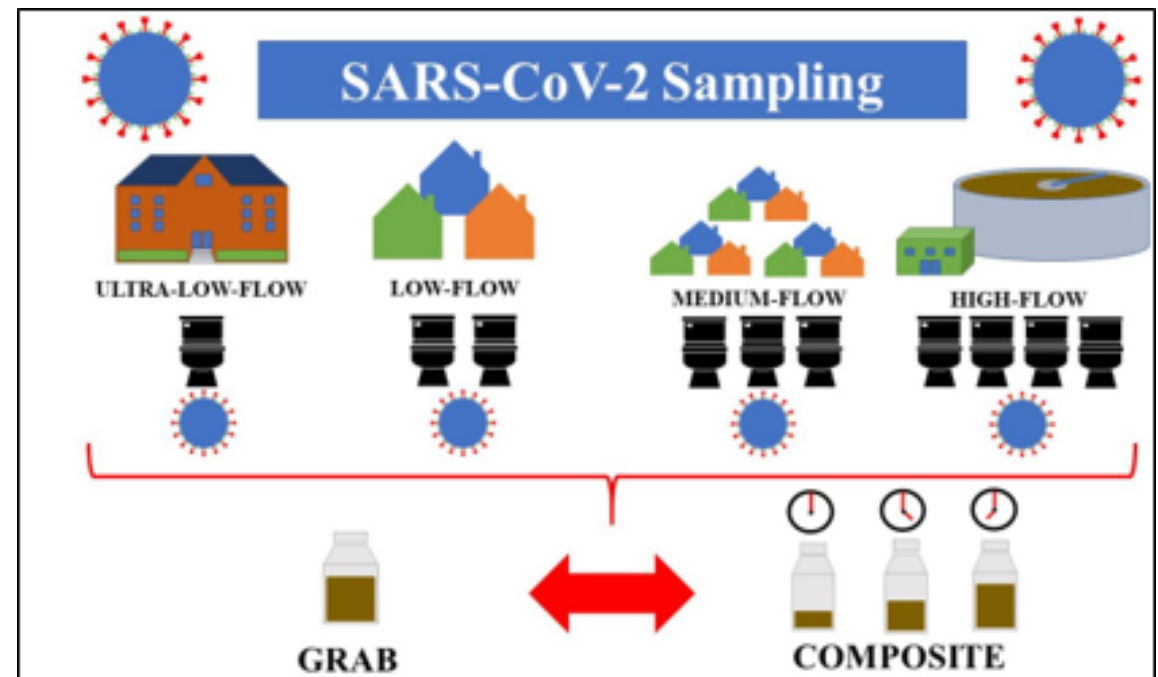
Wastewater Detection Precedes Clinical Cases

- Representative National Wastewater Surveillance System (NWSS) Biobot data



Sampling Strategy

- CDC recommends a “minimum of three samples to detect wastewater trends over time” (Centers for Disease Control and Prevention, 2023).
- High-flow = WWTP
- Medium-flow = neighborhood
- Low-flow = city block



Manual composite sample

Bucket on a rope



Toss into pit



**Retrieve wastewater,
pour into collection
container.**



**Combine
composites, mix,
and aliquot.**



Time weighted composite sample

**ISCO
Autosampler**



**Autosampler hose
enters manhole**



**Wastewater
collection**



Collection carboys



Grab sample

Manhole



Dry manhole



Todd Hubbard, SHL

Detergents and detritus can affect downstream detection

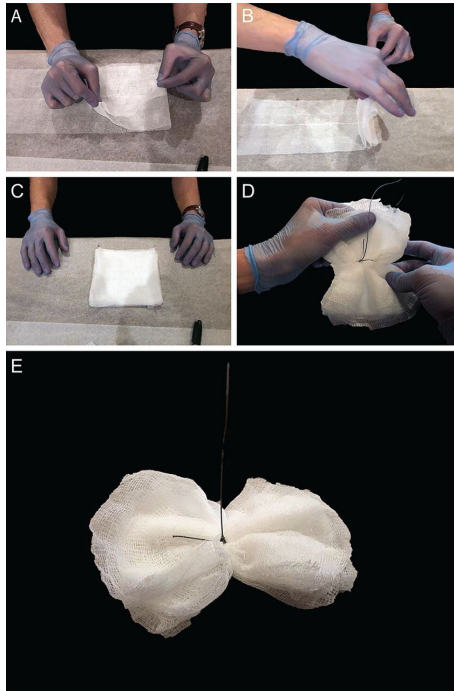


Todd Hubbard, SHL

Bart Everson -
<https://www.flickr.com/photos/editor/48728796337/>

Moore swab sample

Constructing a Moore Swab



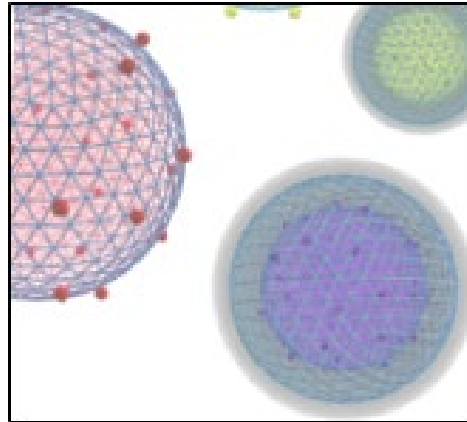
- “A classic environmental surveillance tool involving filtration of flowing surface water and sewage water to recover typhoidal *Salmonella*.”
- Scientific Figure on ResearchGate. Available from: https://www.researchgate.net/figure/Constructing-a-Moore-swab-A-and-B-A-length-of-gauze-6-inches-by-48-inches-is-folded_fig2_340900006 [accessed 24 Mar 2025]
- Sikorski, Michael & Levine, Myron. (2020). Reviving the “Moore swab”: a classic environmental surveillance tool involving filtration of flowing surface water and sewage water to recover typhoidal *Salmonella*. *Applied and Environmental Microbiology*. 86. 10.1128/AEM.00060-20.

Current wastewater test methods

Processing



Concentration



Ceres Nanotrap Particles

Extraction



Amplification

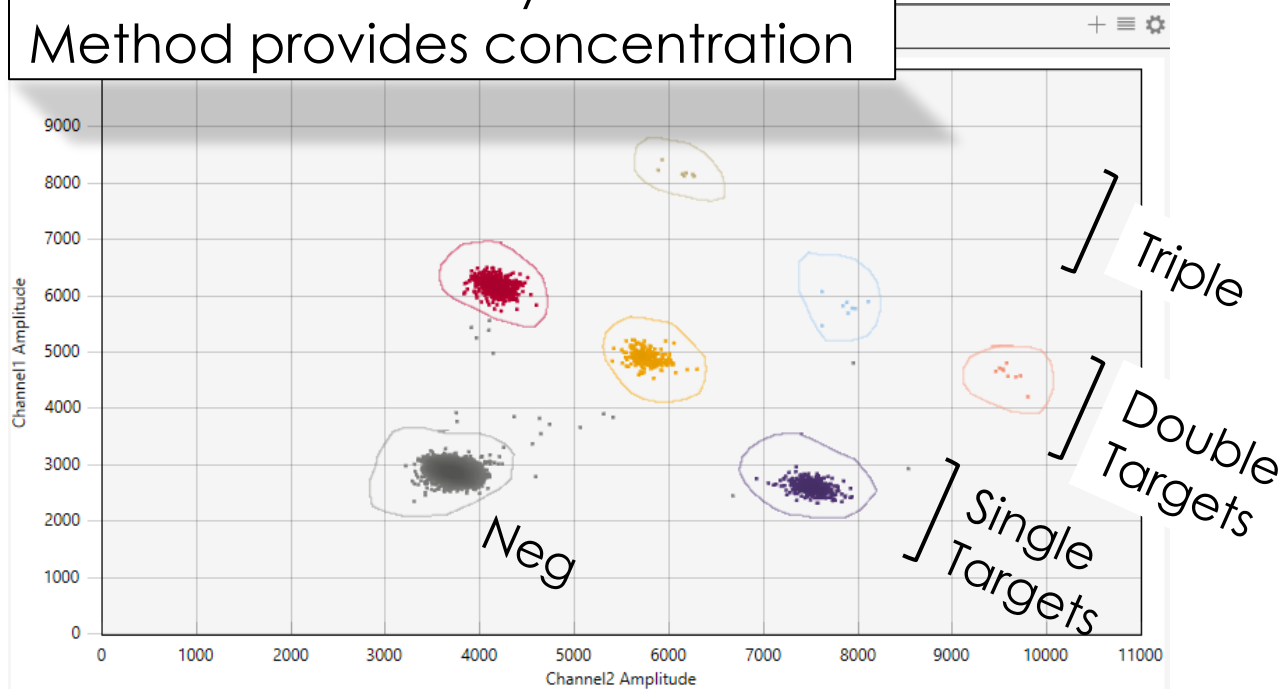


Droplet digital PCR and digital PCR

- Improved sensitivity
- Quantitative result for targets

Droplet Digital PCR Analysis

Bio-Rad PREvalence Assay
3 targets = 2 dyes
Full Validation Study
Method provides concentration



Two COVID-19 gene targets:

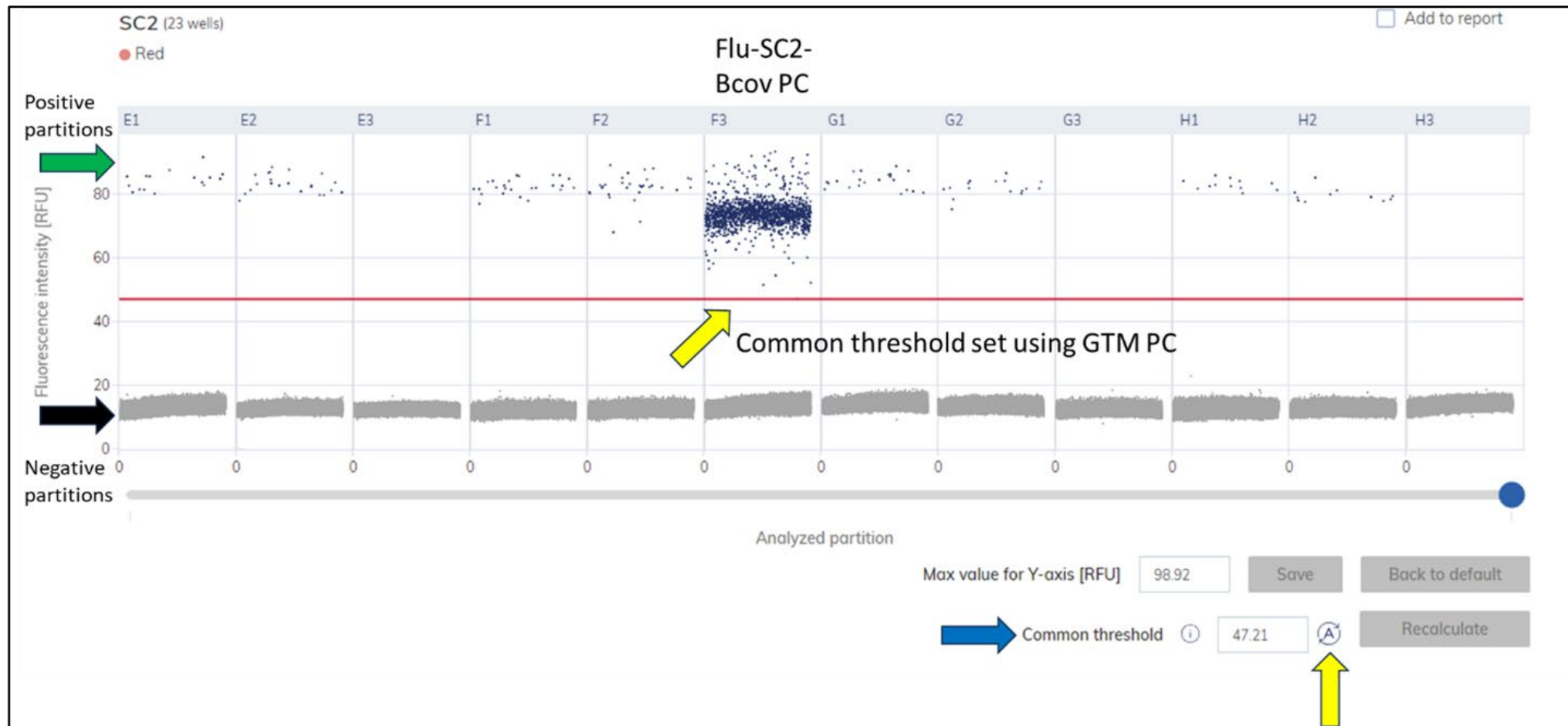
- N2 gene
- E gene

Spike amplification target:

- MHV (murine hepatitis virus)

Results reported through SHL LIMS system to publicly accessible dashboard

Digital PCR Analysis



Wastewater Reports

- Wastewater Sequencing
- Wastewater PCR
 - Qualitative Result
 - Pathogen target
 - Quantitative Result
 - Pathogen target
 - Fecal indicator
 - Normalized Result

Sample
info

 STATE HYGIENIC LABORATORY		ANALYTICAL REPORT		1-800-421-IOWA (4692)
Collection Location	Collector and Phone	Client Reference	Accession # 2517426	
IA	Collected 2024-10-30 13:00	Received 2024-10-31 10:00	Project covid ww wwtp	
Report To			Sample Description wastewater	
			Sample Type Non-Drinking Water	
			Sample Source	
			Sample Note(s) 1	
ADDITIONAL SAMPLE INFORMATION				
Flow Rate (mgd): 0.661 Zip Code: NWSS Sample Location: wwtp NWSS Sample Location Specify: Not Applicable			Institution Type: not institution specific WWTP Name: NWSS Sample Type: 24-hr time-weighted composite NWSS Sample Matrix: raw wastewater	

Results

RESULTS OF ANALYSIS - FINAL REPORT		
TEST	RESULT	ANALYSIS NOTE(S)
Wastewater SARS-CoV-2, Next Generation Sequencing		
Most Abundant Lineage	KP.3.1.1	
Lineage %	41.0	
Coverage Breadth %	90.4	
QC pass	Yes	
SARS Flu RSV in Wastewater, Digital PCR		
Influenza A	Not Detected	2
Influenza A (M1 gene) copies/L	0	
Influenza A Normalized to Fecal Material	0	
Influenza B	Not Detected	
Influenza B (NS1 gene) copies/L	0	
Influenza B Normalized to Fecal Material	0	
SARS-CoV-2	Detected	
SARS-CoV-2 (N gene) copies/L	59000	
SARS-CoV-2 Normalized to Fecal Material	0.00019	
RSV	Not Detected	
RSV (L gene) copies/L	0	
RSV Normalized to Fecal Material	0	
Fecal Material (PMMoV) copies/L	310000000	

Notes

SAMPLE AND ANALYSIS NOTES	
1. Unless otherwise noted, the sample met container and preservation requirements for the analysis requested. Please review carefully your sample results for additional analyte comments or method exceptions.	
2. SHL Wastewater Surveillance link: https://covidtesting.shl.uiowa.edu/wastewater-surveillance-covid-19	
Normally, a quantitative result (copies/L) for each analyte would not be reported if the value is below the Limit of Quantitation (LoQ) because, at very low levels of analyte, precision is reduced. However, to analyze trends, values are reported if they are above the Limit of Blank (LoB). LoB is the highest apparent analyte concentration expected to be found when samples containing no analyte were tested.	

Interpreting wastewater PCR results

Test	Result	Units	Definition
SARS-CoV-2	Detected	Not applicable	SARS-CoV-2 (N gene) value >= Limit of Detection (LoD)
SARS-CoV-2 (N gene)	120000	copies/liter	PCR Gene Target
SARS-CoV-2 Normalized to Fecal Material	0.0014	Unitless	The mean of SARS-CoV-2, or Influenza A/B, or RSV in copies/L wastewater, divided by the amount of the fecal matter indicator virus (pepper mild mottle virus) in copies/L wastewater.
Fecal Material (PMMoV)	310000000	copies/liter	Amount of pepper mild mottle virus in wastewater sample in copies/L wastewater

Objective



Hypothesis

Null hypothesis: $\beta = 0$

Increasing wastewater sample collections from one sample to two samples per week will have no effect in predicting hospitalizations. (Slope = 0)

Alternative hypothesis: $\beta \neq 0$

Increasing sample collections from one sample to two samples per week will predict hospitalizations. (Slope $\neq 0$)

Sampling Frequency and Hospitalization Trends

Simple Linear Regression Model

$$y = \alpha + \beta x + \epsilon$$

Fitted Simple Linear Regression Line

$$Y = a + bx$$

$$\text{Hospitalization} = \alpha + \log_{10} \text{normalized SARS-CoV-2 virus} \times \beta$$

Methods



Test Methods

- January 2023 – November 2023
 - Concentration: Ceres Nanotrap Particles
 - Instrument: Bio-Rad QXOne
 - Assay: Bio-Rad PREvalence ddPCR SARS-COV-2 Wastewater Quantification Kit
 - Limit-of-Detection = 11,000 copies/L
- December 2023 – April 2024
 - Concentration: Ceres Nanotrap Particles
 - Instrument: QIAcuity 4
 - Assay: GT Molecular GT-Digital Influenza and SARS-Cov-2 Wastewater Surveillance Multiplexed Assay Kit
 - Limit-of-Detection = 10,000 copies/L

Wastewater Samples

- 24-hour composite wastewater samples
- Davenport Wastewater Treatment Plant
- Collection dates
 - One wastewater sample per week
 - January 9, 2023 – September 18, 2023 (n = 39)
 - One wastewater sample
 - September 25, 2023 – April 8, 2024 (n = 51)

Hospitalization Data

- Inpatients at Genesis Healthcare in Davenport, Iowa with a positive COVID-19 PCR test from January 1, 2023 – April 22, 2024 (n = 2440)
 - No patient information collected
 - Date of PCR positive
 - Number of inpatients

Results



Hospitalizations Statistics

7-day rolling average of daily PCR positive COVID-19 inpatients

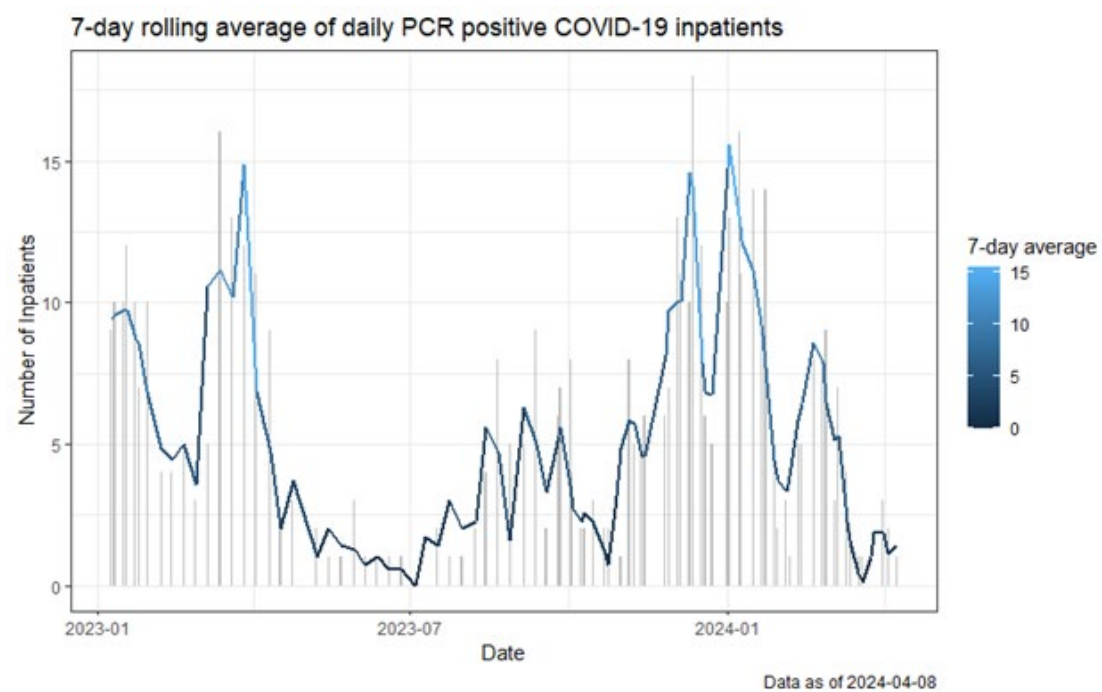


Figure 1 Graph of daily hospitalized patients with the 7-day rolling average overlay.

14-day rolling average of daily PCR positive COVID-19 inpatients

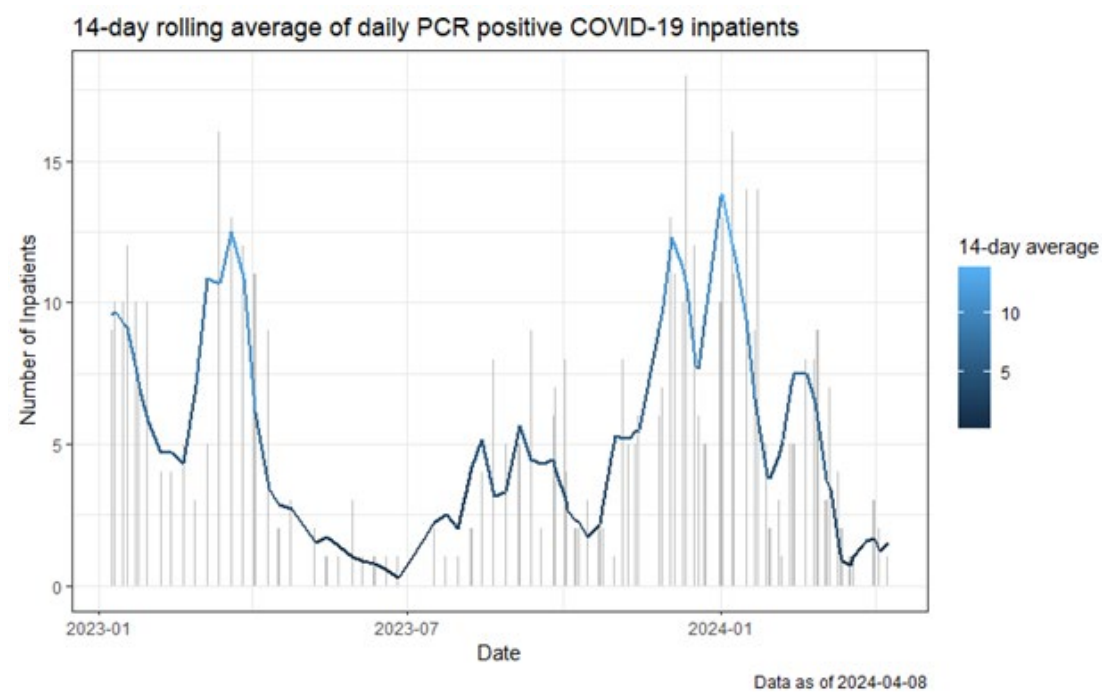


Figure 2 Graph of daily hospitalization patients with the 14-day rolling average overlay.

Hospitalizations by sample(s) per week

One sample/week N=39

7-day Hospitalizations	
Mean (SD)	4.9 (3.7)
Median (IQR)	4.4 (1.6, 7.7)
Range	0.0, 14.9

14-day Hospitalizations	
Mean (SD)	4.8 (3.7)
Median (IQR)	4.3 (1.9, 7.0)
Range	0.3, 12.5

Two samples/week N=51

7-day Hospitalizations	
Mean (SD)	5.8 (4.1)
Median (IQR)	5.3 (2.4, 8.0)
Range	0.1, 15.6

14-day Hospitalizations	
Mean (SD)	5.7 (3.7)
Median (IQR)	5.2 (2.5, 7.7)
Range	0.7, 13.9

Normalized wastewater by sample(s) per week

One sample/week N=39

Normalized SARS-CoV-2	
Mean	0.00050
Median	0.00058
Range	0.00000, 0.00110

Normalized SARS-CoV-2 x10,000	
Mean (SD)	5.0 (3.7)
Median (IQR)	5.8 (1.3, 8.6)
Range	0.0, 11.0

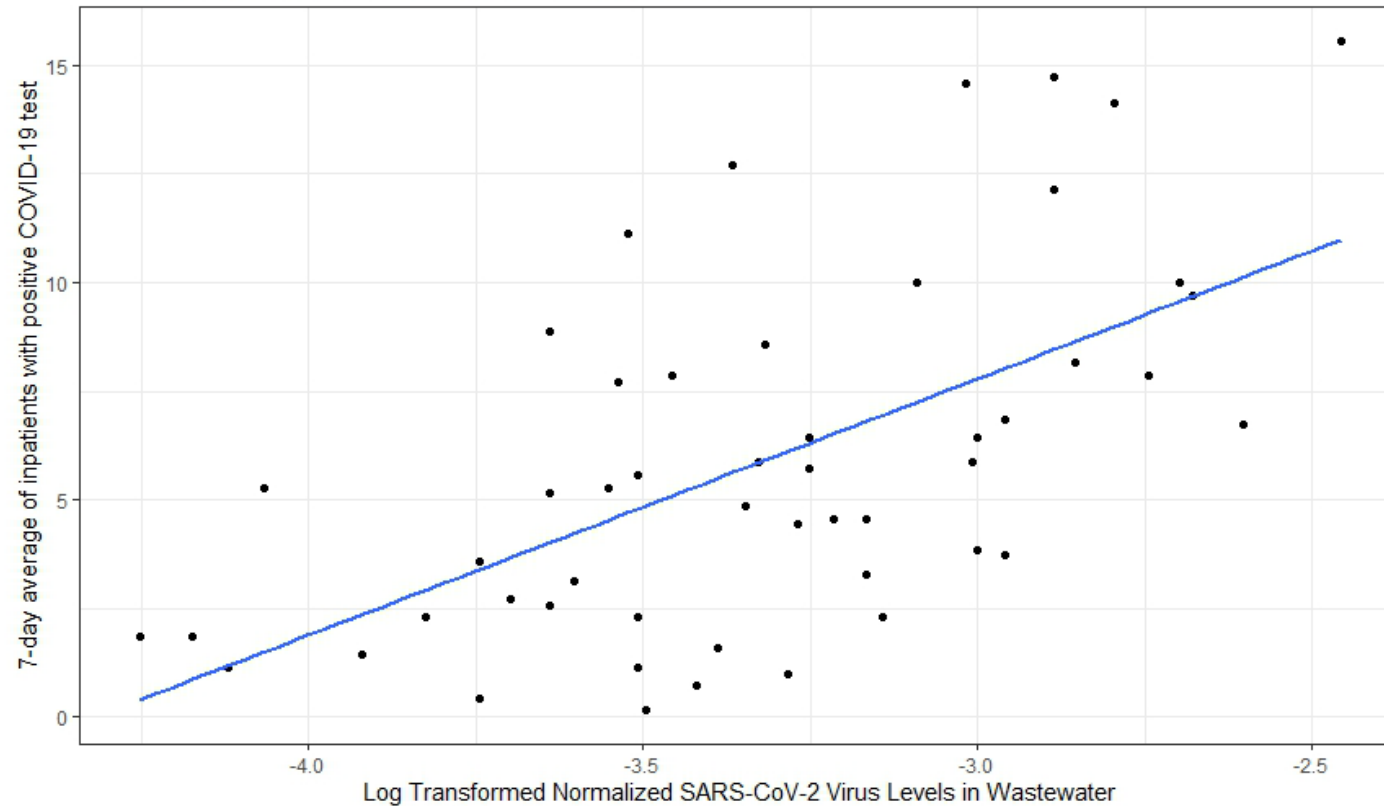
Two samples/week N=51

Normalized SARS-CoV-2	
Mean	0.00072
Median	0.00047
Range	0.00006, 0.00350

Normalized SARS-CoV-2 x10,000	
Mean (SD)	7.2 (7.0)
Median (IQR)	4.7 (2.5, 10.0)
Range	0.6, 35.0

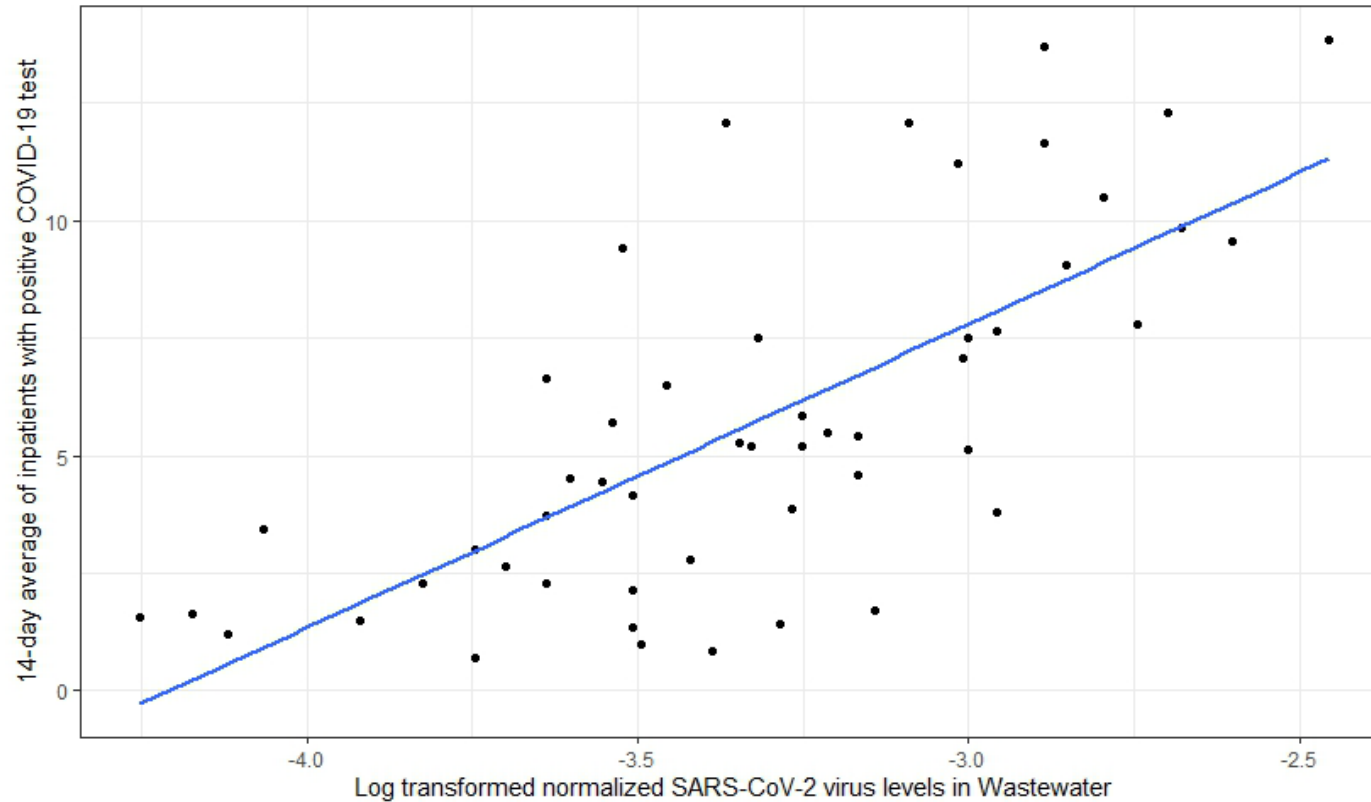
7-day, two-sample, linear regression model

Figure 1. Inpatients with COVID-19 positive test against normalized SARS-CoV-2 levels in wastewater.



14-day, two-sample, linear regression model

Figure 2. Inpatients with COVID-19 positive test against normalized SARS-CoV-2 levels in wastewater.



7-day, regression coefficients & test statistics

Model	F	Df	P-value	R ² adjusted
7-day inpatients, log10 normalized wastewater levels	27.63	1, 49	< 0.001	0.35

Coefficients	Estimate	Standard Error	P-value
7-day inpatients	25.50	3.76	< 0.001
Log10 Wastewater	5.90	1.12	<0.001

14-day, regression coefficients & test statistics

Model	F	Df	P-value	R ² adjusted
14-day inpatients, log10 normalized wastewater levels	52.61	1, 49	< 0.001	0.51

Coefficients	Estimate	Standard Error	P-value
14-day inpatients	27.20	2.99	< 0.001
Log10 Wastewater	6.46	0.89	<0.001

Discussion



Summary

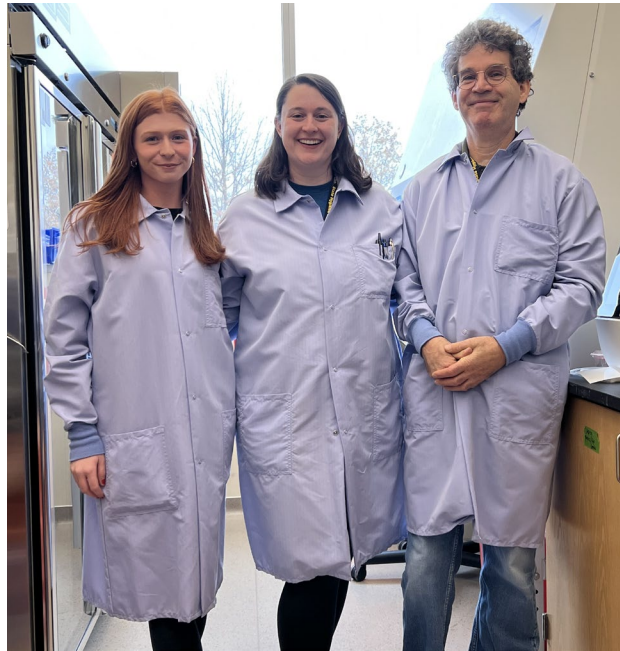
- Based on the analysis of hospitalizations for the largest healthcare system in Scott County, Iowa there is a significant relationship between SARS-CoV-2 wastewater levels predicting hospitalization rates at 7-days and 14-days when sample collection is increased to two samples per week.
- Hospitalization numbers only include inpatients with a positive COVID-19 PCR test and do not include total inpatients per day.
- Screening of all inpatients for COVID-19 ceased on January 1st, 2023, so testing for COVID-19 was ordered because symptoms were present that indicated COVID-19 infection.
- Multiplying normalized SARS-CoV-2 wastewater levels, log transforming, improved normality and could be included in future studies with the goal of improving model fit.
 - Shapiro-Wilk test was performed using model residuals
 - Normality was rejected for every one-sample and two-sample regression models except for the two sample/week, 14-day hospitalizations regression model.
- Thank you to Dr. Cynthia Cass and Dr. Michael Pentella.
- Thank you to the Association of Public Health Laboratories.

Iowa Wastewater Team

Limnology Wastewater Collection



Environmental Microbiology Wastewater dPCR/ddPCR



Molecular Biology Sequencing/Bioinformatics



References

Association of Public Health Laboratories (Ed.). (2022). *SARS-CoV-2 Wastewater Surveillance Testing Guide for Public Health Laboratories*. Association of Public Health Laboratories. <https://www.aphl.org/aboutAPHL/publications/Documents/EH-2022-SARSCoV2-Wastewater-Surveillance-Testing-Guide.pdf>

Centers for Disease Control and Prevention. (2023, March 29). *Developing a Wastewater Surveillance Sampling Strategy* [Government]. Centers for Disease Control and Prevention. <https://www.cdc.gov/nwss/sampling.html>

Kilaru, P., Hill, D., Anderson, K., Collins, M. B., Green, H., Kmush, B. L., & Larsen, D. A. (2023). Wastewater Surveillance for Infectious Disease: A Systematic Review. *American Journal of Epidemiology*, 192(2), 305–322. <https://doi.org/10.1093/aje/kwac175>

Kirby, A. E., Walters, M. S., Jennings, W. C., Fugitt, R., LaCross, N., Mattioli, M., Marsh, Z. A., Roberts, V. A., Mercante, J. W., Yoder, J., & Hill, V. R. (2021). Using Wastewater Surveillance Data to Support the COVID-19 Response—United States, 2020-2021. *MMWR. Morbidity and Mortality Weekly Report*, 70(36), 1242–1244. <https://doi.org/10.15585/mmwr.mm7036a2>

Parkins, M. D., Lee, B. E., Acosta, N., Bautista, M., Hubert, C. R. J., Hrudehy, S. E., Frankowski, K., & Pang, X.-L. (2024). Wastewater-based surveillance as a tool for public health action: SARS-CoV-2 and beyond. *Clinical Microbiology Reviews*, 37(1), e0010322. <https://doi.org/10.1128/cmr.00103-22>

Tiwari, A., Ahmed, W., Oikarinen, S., Sherchan, S. P., Heikinheimo, A., Jiang, G., Simpson, S. L., Greaves, J., & Bivins, A. (2022). Application of digital PCR for public health-related water quality monitoring. *The Science of the Total Environment*, 837, 155663. <https://doi.org/10.1016/j.scitotenv.2022.155663>

Galani A, Aalizadeh R, Kostakis M, Markou A, Alygizakis N, Lytras T, Adamopoulos PG, Peccia J, Thompson DC, Kontou A, Karagiannidis A, Lianidou ES, Avgeris M, Paraskevis D, Tsiodras S, Scorilas A, Vasiliou V, Dimopoulos MA, Thomaidis NS. SARS-CoV-2 wastewater surveillance data can predict hospitalizations and ICU admissions. *Sci Total Environ*. 2022 Jan 15;804:150151. doi: 10.1016/j.scitotenv.2021.150151. Epub 2021 Sep 7. PMID: 34623953; PMCID: PMC8421077.

Sikorski, Michael & Levine, Myron. (2020). Reviving the “Moore swab”: a classic environmental surveillance tool involving filtration of flowing surface water and sewage water to recover typhoidal *Salmonella*. *Applied and Environmental Microbiology*. 86. 10.1128/AEM.00060-20.

IOWA

State Hygienic Laboratory

Thank you

→ uiowa.edu

Ryan Jepson
Environmental Microbiology
Manager
State Hygienic Laboratory at the
University of Iowa

319-335-4269
ryan-jepson@uiowa.edu

IOWA